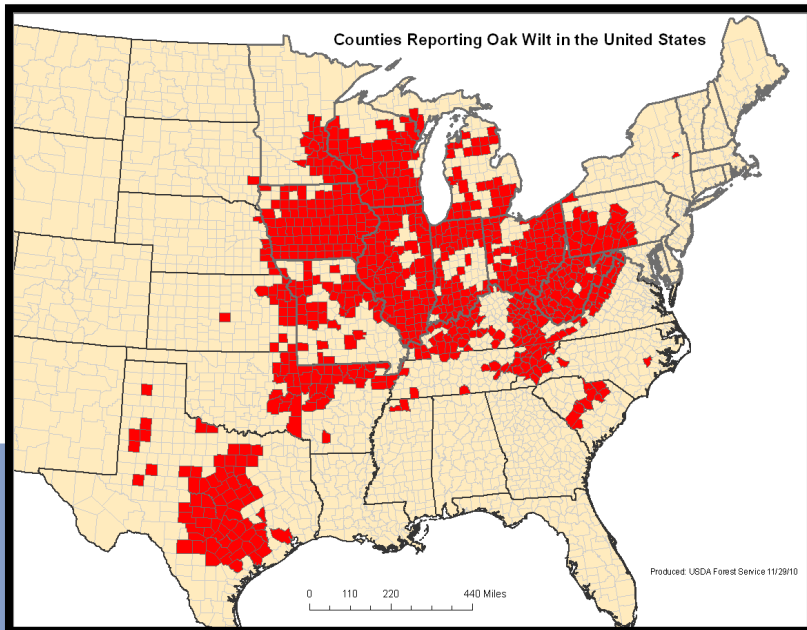


IMPROVING OAK WILT DIAGNOSTICS IN MINNESOTA

Anna Yang
Department of Plant Pathology
University of Minnesota

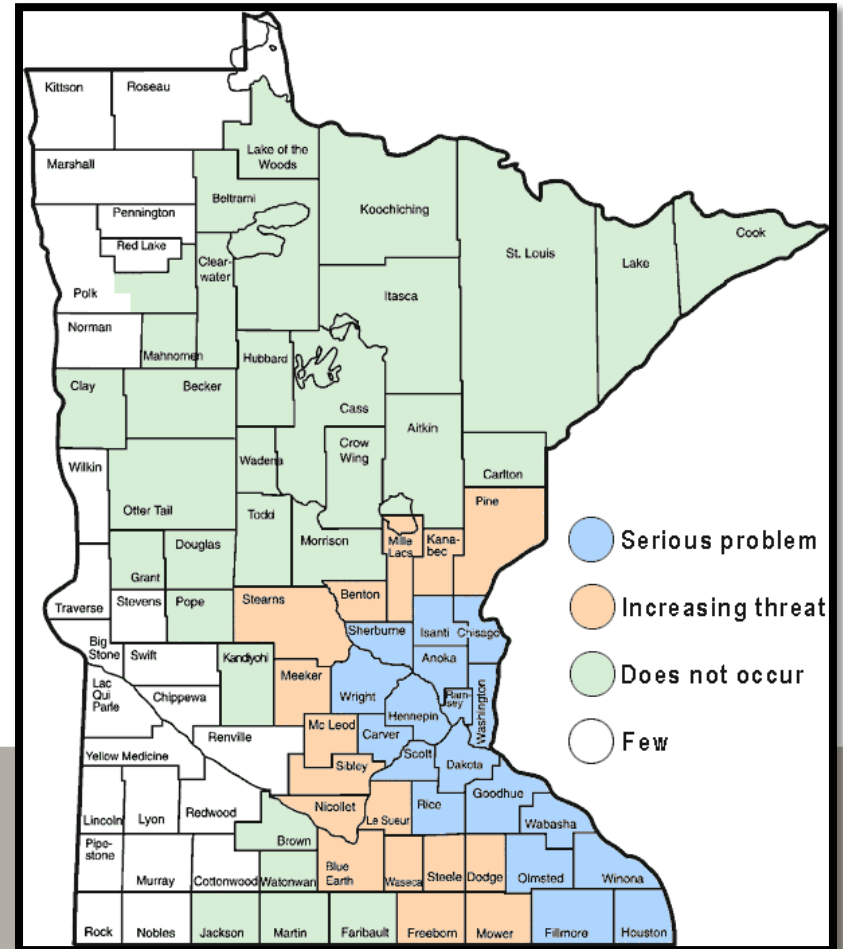
OAK WILT

- Significant disease of oak species
- Caused by *Ceratocystis fagacearum*



U.S. Distribution in 2010

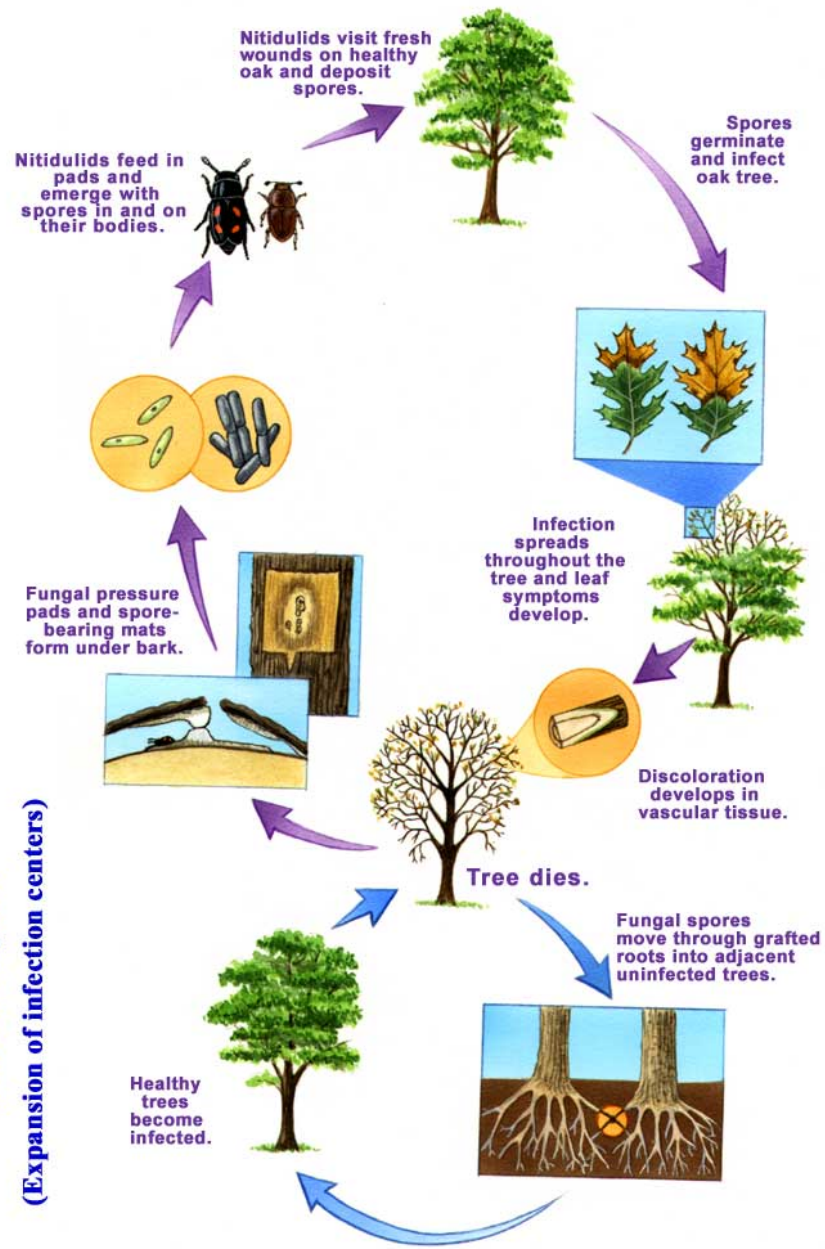
Source: Q. Chavez and J. Pokorny. Northeastern Area State and Private Forestry, U.S. Forest Service.



MN Distribution

Source: J. Juzwik and D. French, 2002.

Overland Spread (Initiation of new infection centers)



**Root Graft Spread
(Expansion of infection centers)**

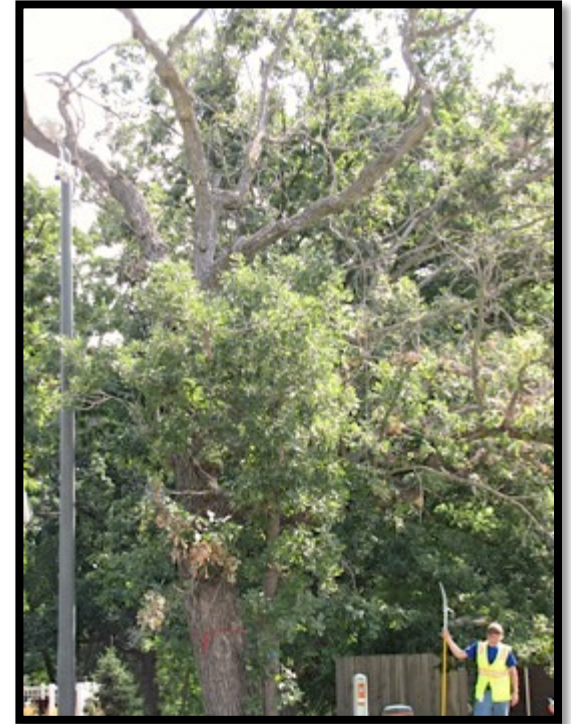
SYMPTOMS



Red Oak



Bur Oak



White Oak

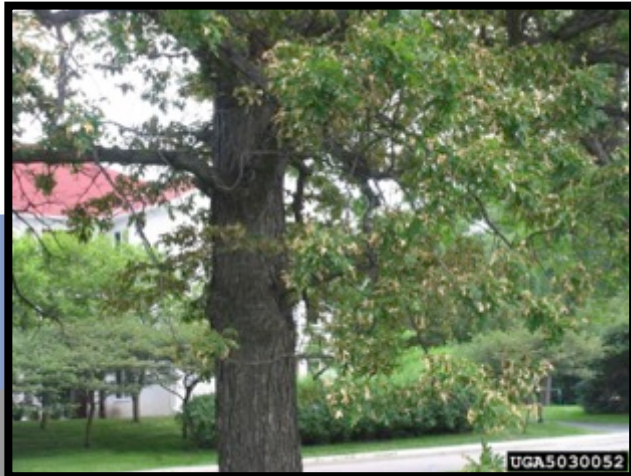
Other insects and disease may mimic symptoms

COPY CATS



Bur Oak Blight

Source: T. Harrington, Iowa State University



Anthrachnose

Source: J. O'Brien, U.S. Forest Service



Twolined Chestnut Borer

Source: S. Katovich, U.S. Forest Service

And many more!

DISEASE MANAGEMENT

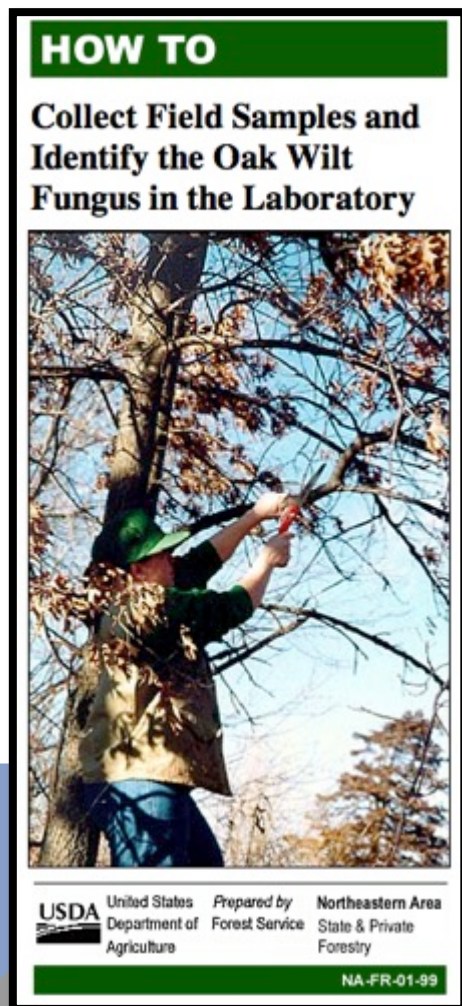
- Effective management options available
- Dependent on *accurate* and *timely* diagnosis



Vibratory Plow Lines

Source: B. Cook, Michigan State University

CURRENT DIAGNOSTICS



Approach

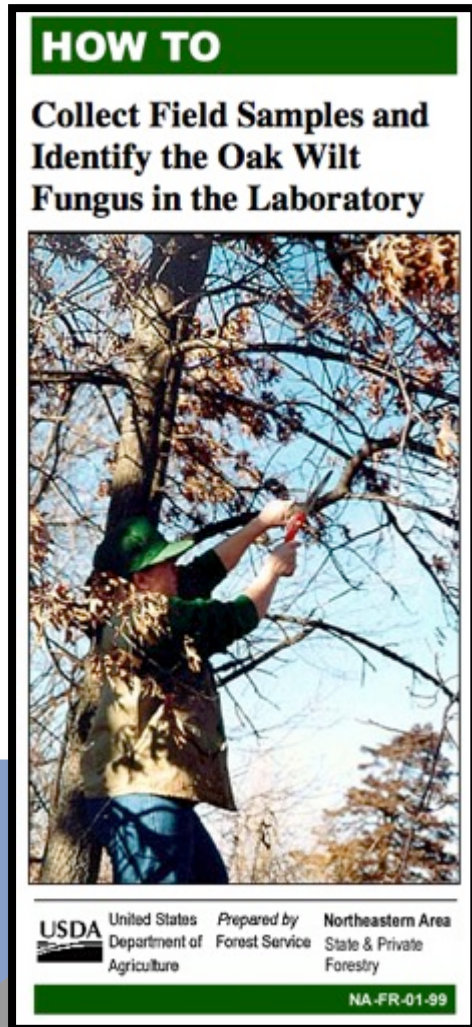
- Visual assessment
- Sampling for diagnostic clinic

Lab Diagnosis

- Isolation from wood chips
- Long incubation
- Dependent on sample quality
- Occurrence of false negatives

Management / Treatment

CURRENT DIAGNOSTICS



Approach

- Visual assessment
- Sampling for diagnostic clinic

Sampling Steps:

1. Select partially wilted branch
2. Look for discoloration in sapwood
3. Sample from up to three branches
4. If branches are too high, sample main stem
5. Keep samples cool during transport

CURRENT DIAGNOSTICS



Lab Diagnosis

- Isolation from wood chips
- Long incubation
- Dependent on sample quality
- Occurrence of false negatives

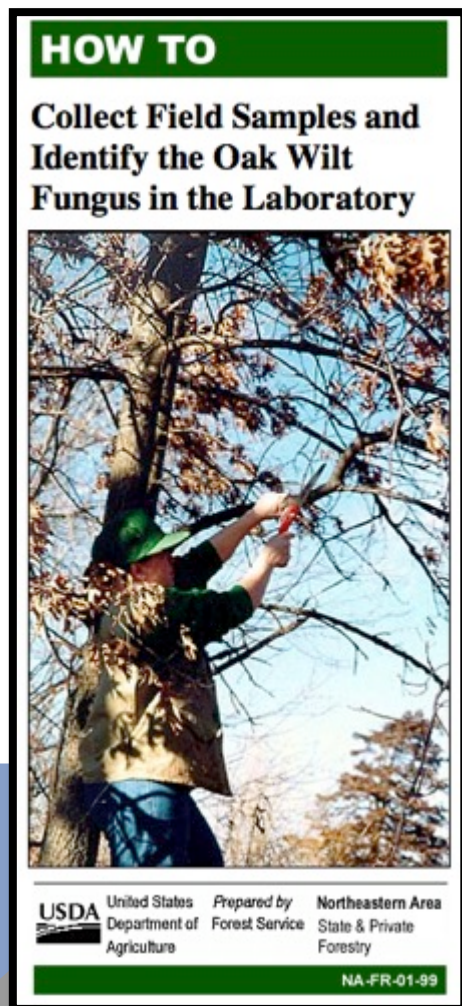


Poor Sample Quality:

- Dead branches
- Dry samples
- Other fungi present

Total Time: 6-14 days

CURRENT DIAGNOSTICS



Approach

- Visual assessment
- Sampling for diagnostic clinic

Lab Diagnosis

- Isolation from wood chips
- Long incubation
- Dependent on sample quality
- Occurrence of false negatives

Effective treatment depends
on early and accurate
diagnosis

NEW METHODS

Rapid and accurate detection of *Ceratocystis fagacearum* from stained wood and soil by nested and real-time PCR

By C. P. Wu^{1,2}, G. Y. Chen¹, B. Li², H. Su², Y. L. An², S. Z. Zhen² and J. R. Ye^{1,3}

¹Jiangsu Province Key Laboratory for Prevention and Management of Invasive Species, Nanjing Forest University, Nanjing 210037, China; ²Plant Quarantine Laboratory, Jiangsu Entry-Exit Inspection and Quarantine Bureau, Nanjing, China; ³E-mail: jrye@njfu.com.cn (for correspondence)

The detection of *Ceratocystis fagacearum* in Texas live oak using real-time polymerase chain reaction

T. KURDYLA (1), D. Appel (1)

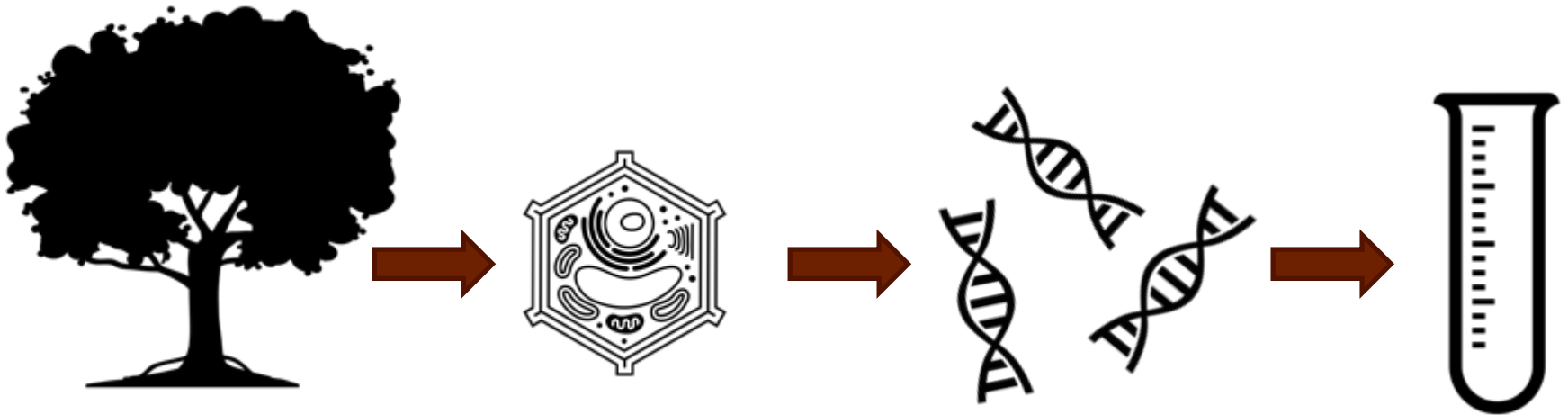
(1) Texas A&M University, College Station, TX, U.S.A.

Phytopathology 101:S95

Developed, but not fully useful yet.

OBTAINING DNA

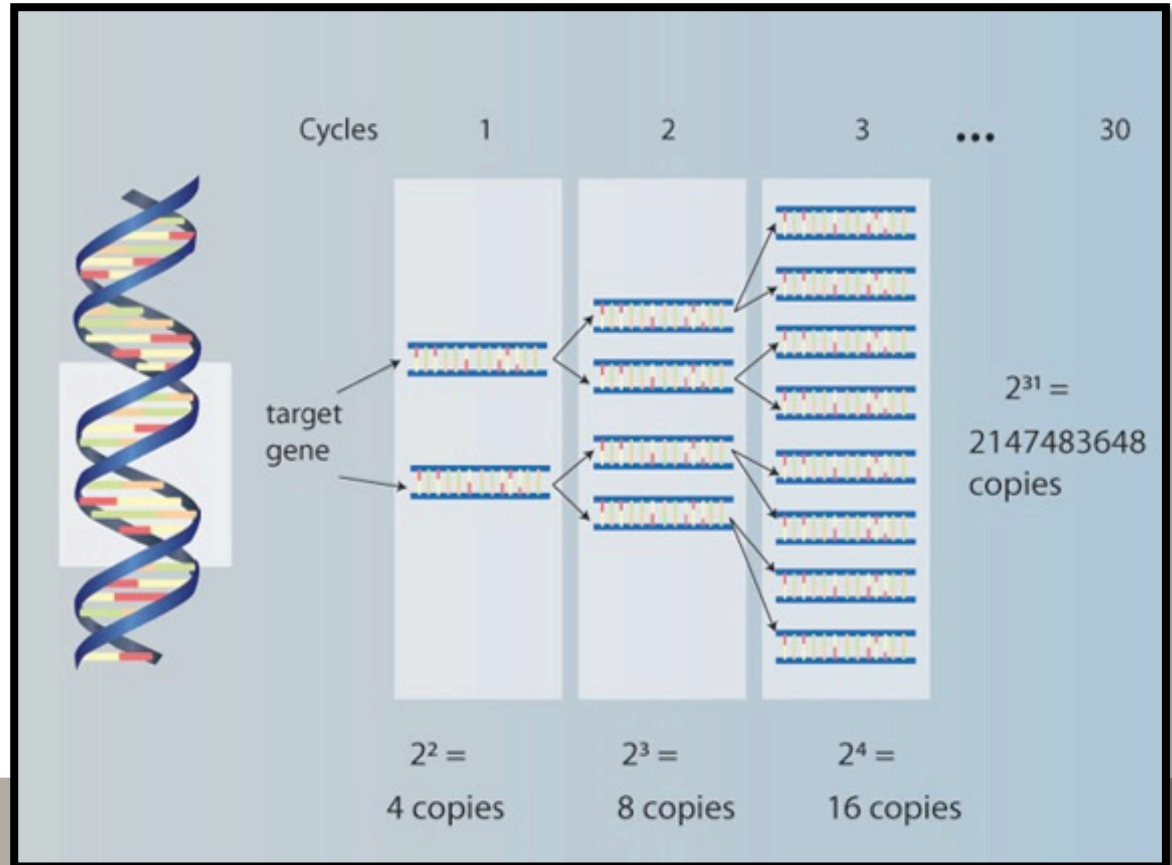
DNA is extracted directly from wood



WHAT IS PCR?

Polymerase Chain Reaction

Making millions of copies of a portion of DNA (think CSI).



Nested PCR

Real-Time PCR

RESEARCH OBJECTIVES

Test and modify nested and real-time PCR protocols for detection of *C. fagacearum* in sapwood




Develop and test field sampling and laboratory processing guidelines



Evaluate reliability and practicality for routine use by diagnostic laboratories



- 1. SAMPLING METHODS**
 - 2. SAMPLE PROCESSING**
 - 3. NESTED PCR + RESULTS**
 - 4. REAL-TIME PCR + RESULTS**
 - 5. CURRENT STATUS**
- 
- A decorative footer consisting of several overlapping geometric shapes. On the left, there is a blue triangle pointing right. Next to it is a grey triangle pointing left. The rest of the bottom area is filled with a solid grey color.

SAMPLING METHODS – YEAR 1

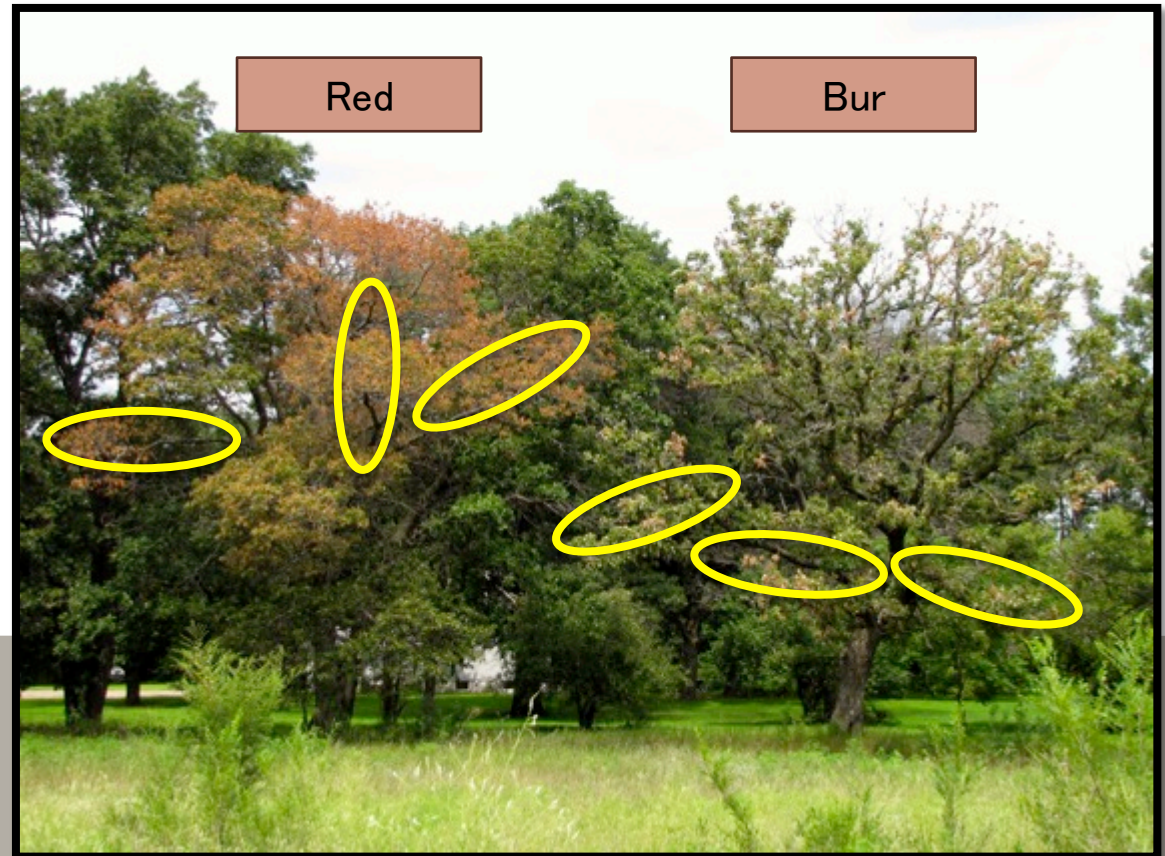
Actively Wilting Crowns

Three branches selected
from each tree:

- Nine red oak
- Eight bur oak
- Eight white oak

Three healthy control
trees of each species

Seven communities
sampled



SAMPLING METHODS – YEAR 2

≥ 1 Year Dead Branches

Three branches selected
from each tree:

- Three bur oak
- Four white oak

Two non-oak wilt
affected control trees for
each species

Seven communities
sampled



SAMPLING METHODS – YEAR 2

≥ 1 Year Dead Red Oak



Three “windows” removed from each red oak tree:

- Six streaking
- Seven mat scar

Two non-oak
wilt killed
control trees

Two
communities
sampled

LABORATORY PROCESSING

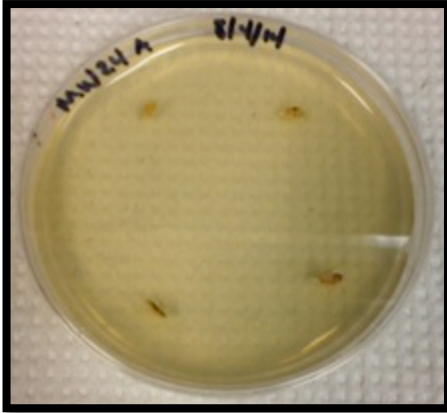


Isolation

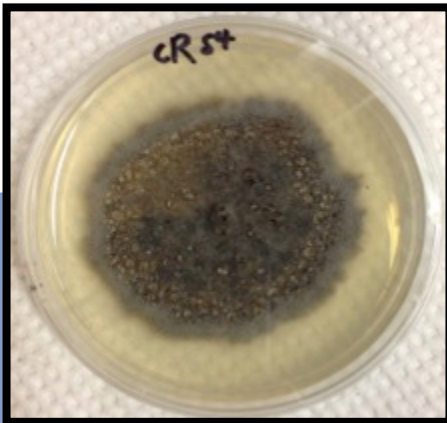
Molecular Detection



Isolation



Plating Wood
Chips



14 day
incubation



Molecular Detection



Sapwood
Drilling



DNA
Extraction

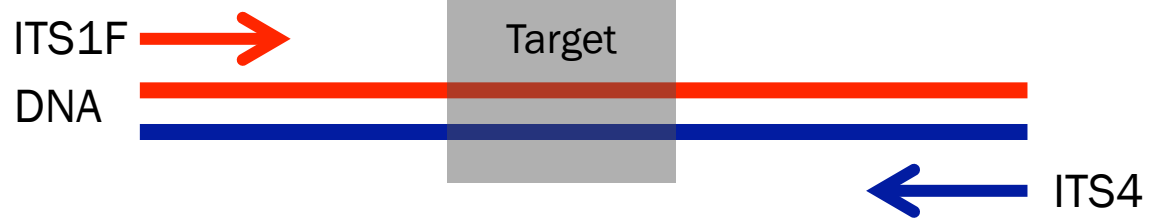
Nested PCR

&

Real-Time PCR

NESTED PCR

Round 1
Amplify all
fungi



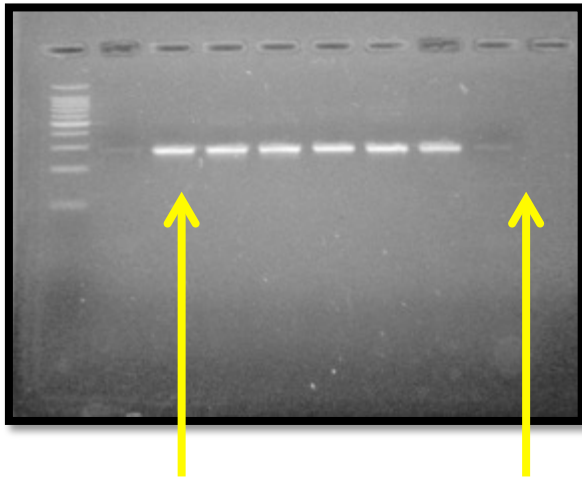
Round 2
Only
amplify C.f.



Final
C.f. DNA
fragments



NESTED PCR



Positive

Negative

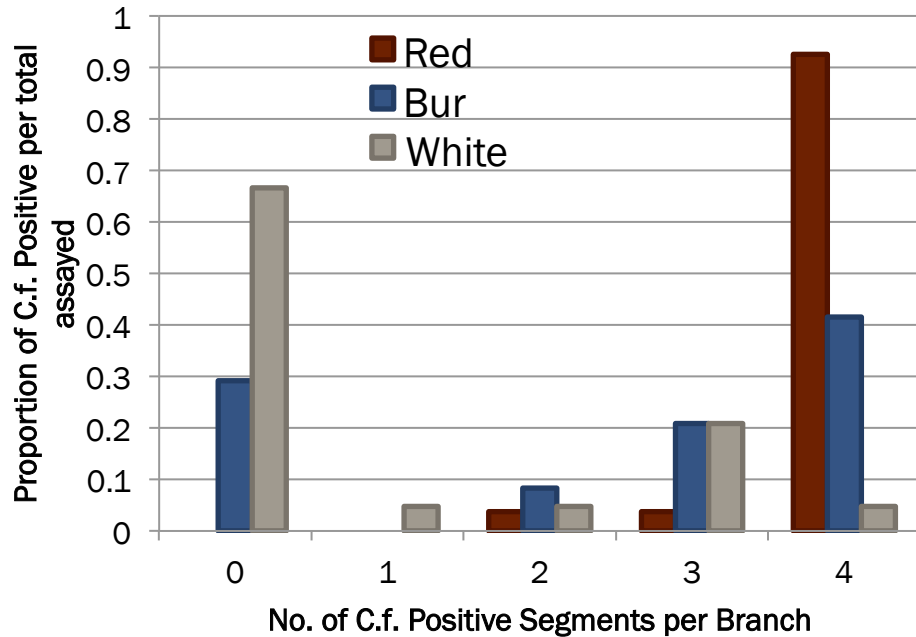
	Description	max score	total score	query cover	E value	Ident	Accession
<input type="checkbox"/>	Ceratocystis fagacearum strain C 660 18S ribosomal RNA gene, partial sequence; internal transcribed spacer	379	379	100%	7e-102	100%	KC305153.1
<input type="checkbox"/>	Ceratocystis fagacearum strain C 520 18S ribosomal RNA gene, partial sequence; internal transcribed spacer	379	379	100%	7e-102	100%	KC305152.1

Extraction: 2 hours
Nested PCR : 6 hours
Total time: 1-2 days

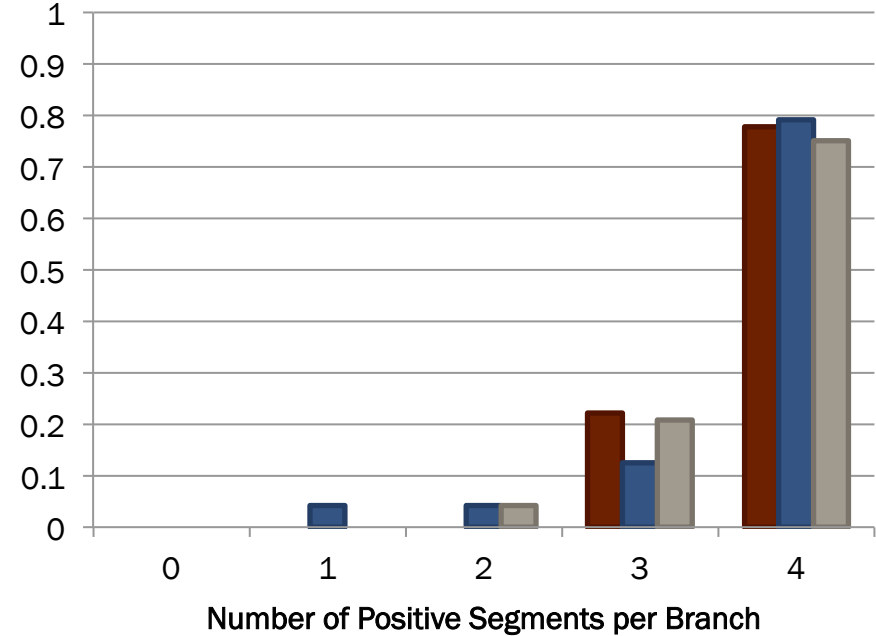
NESTED PCR RESULTS

ACTIVE WILT

Traditional Isolation



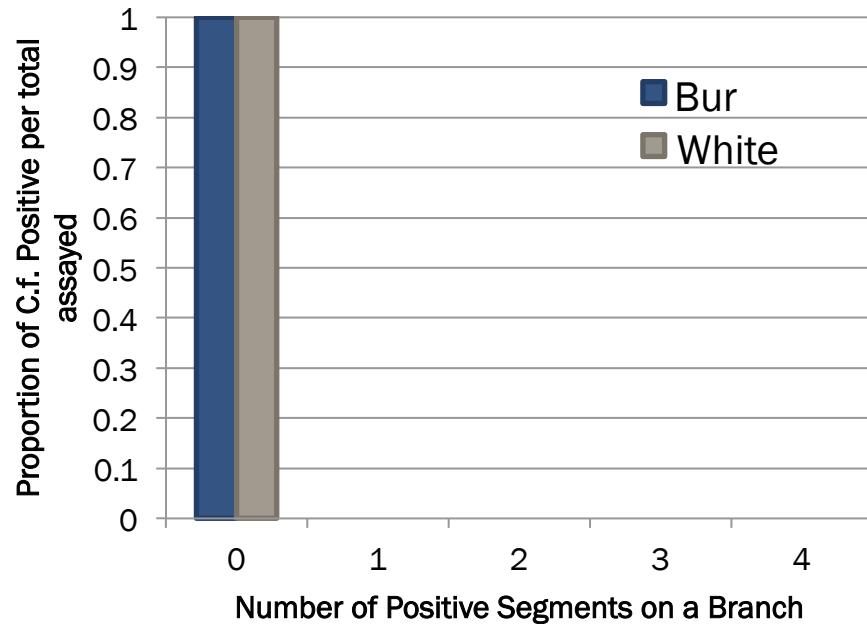
Nested PCR



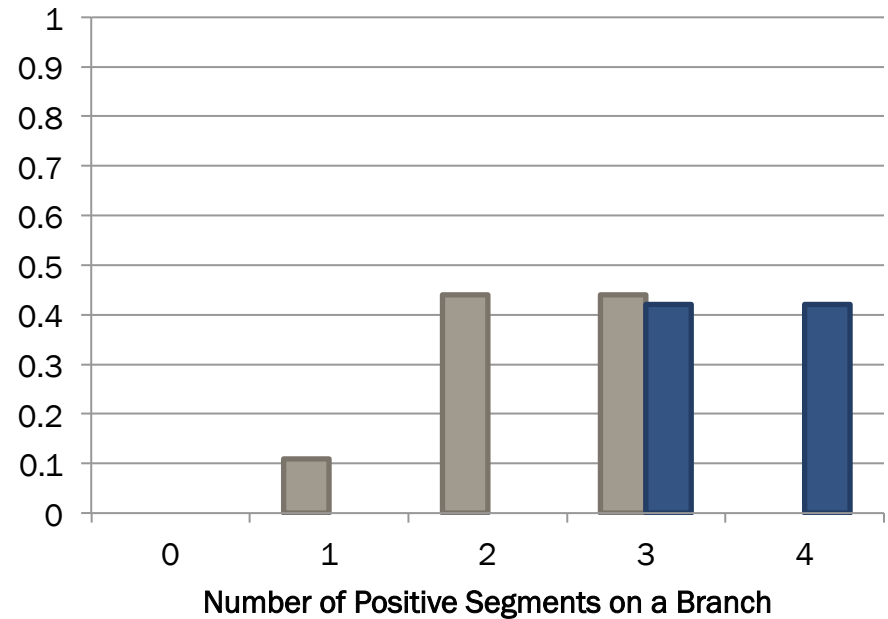
- Red Oak - little difference in detection between methods
- Bur and White Oak - nested PCR superior

≥ 1 YEAR DEAD BRANCHES

Traditional Isolation



Nested PCR

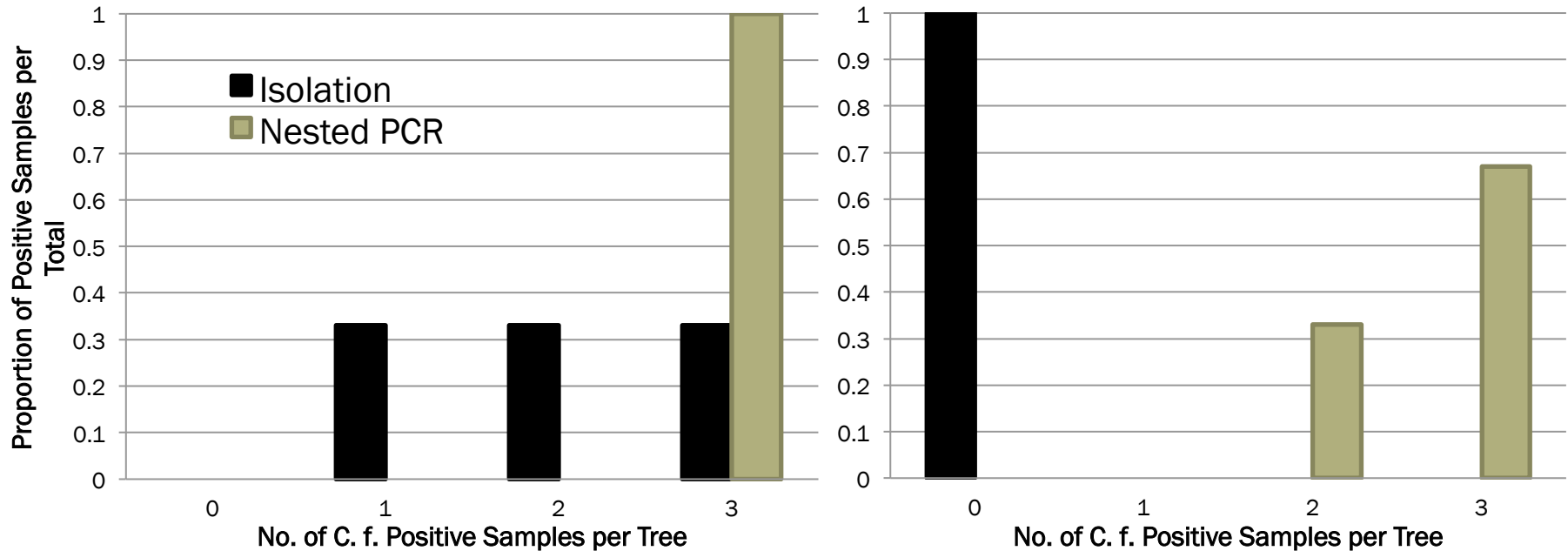


- Bur and White Oak - only detected through nested PCR

RED OAK - MAIN STEM SAMPLES

Streaked Cambium

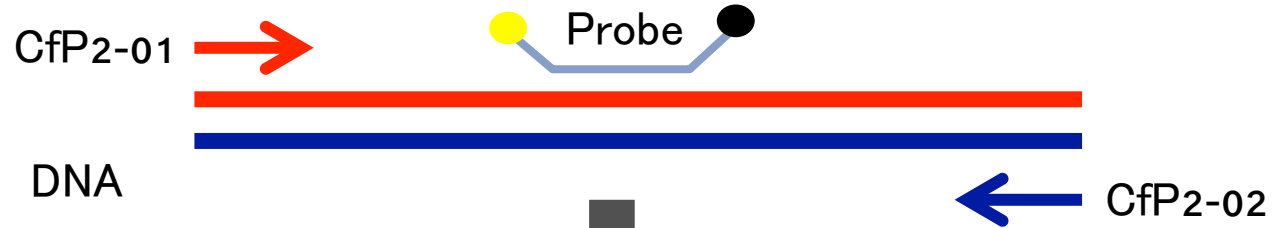
Mat Scars



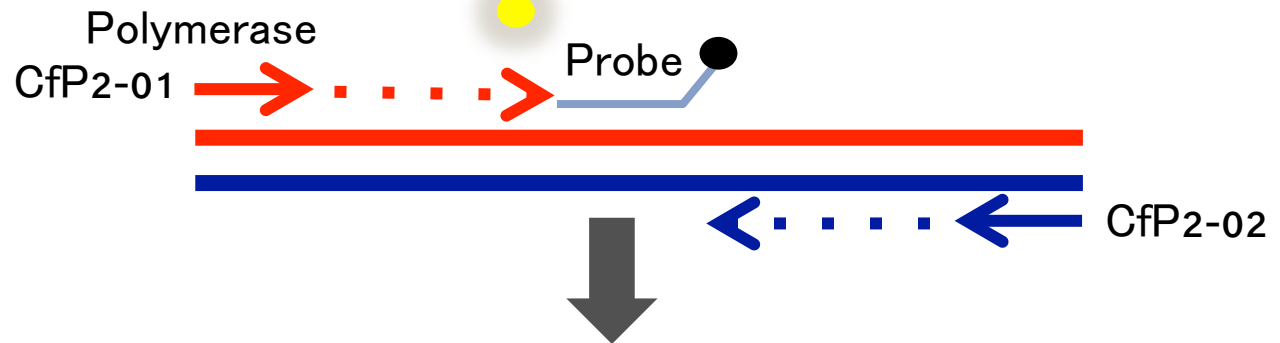
- Streaked Cambium - slightly higher detection through nested PCR
- Mat Scars - only detection through nested PCR

REAL-TIME PCR

Step 1:
Primer +
Probe to
amplify C.f.



Step 2:
“Light up”
every time
DNA is
doubled



Final:
C.f. DNA
and “light
up”
molecules

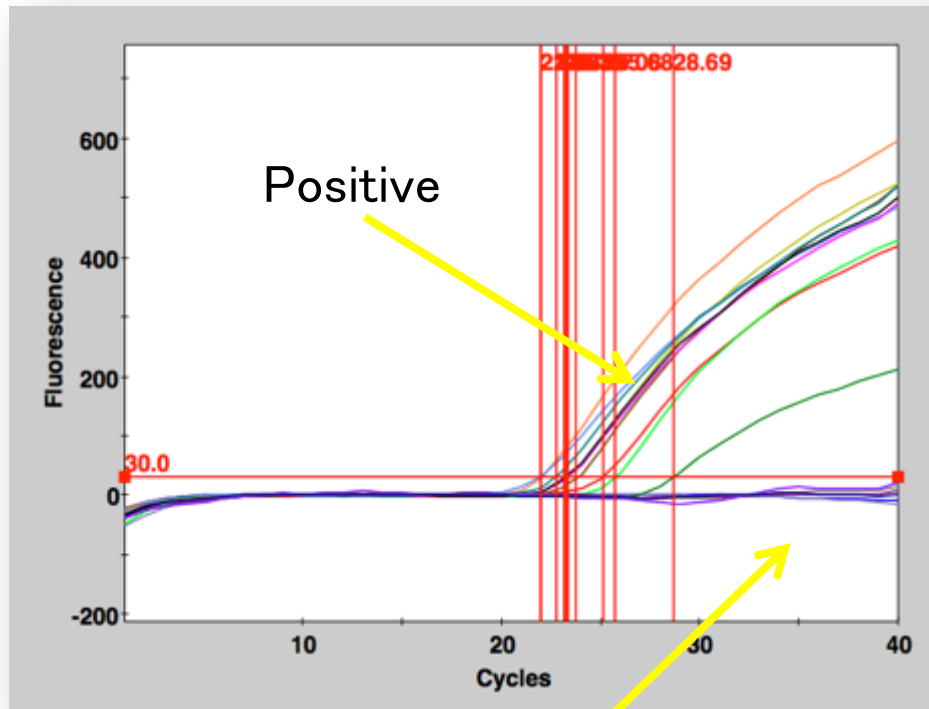


C.f. DNA



Cleavage
Product

REAL-TIME PCR



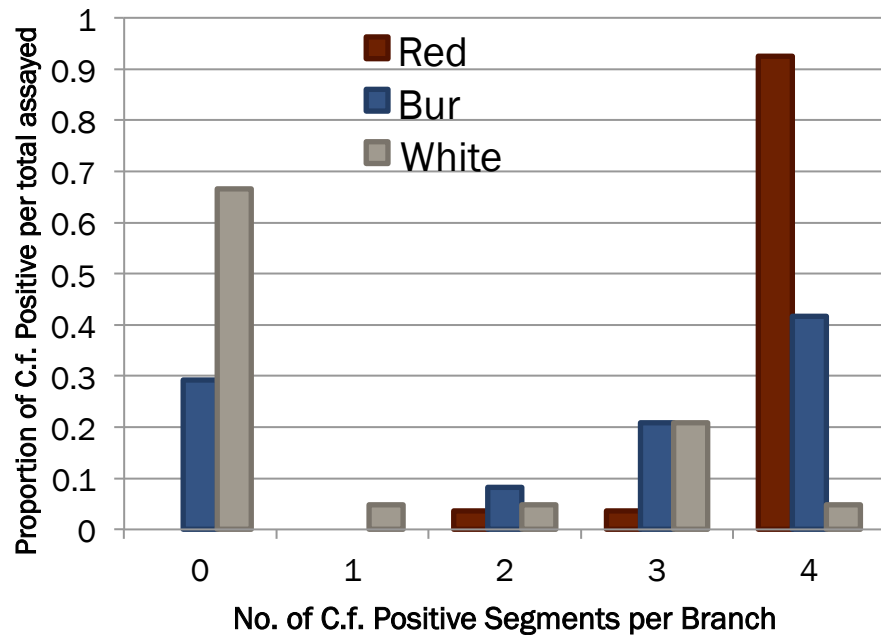
Site ID	Sample ID	FAM Ct	Protocol
A1	NTC	0.00	OW test
A2	NTC	0.00	OW test
A3	CF-0	22.03	OW test
A4	CF-2	28.69	OW test
A5	501	23.33	OW test
A6	501	23.77	OW test
A7	502	25.08	OW test
A8	502	25.68	OW test
A9	503	21.99	OW test
A10	503	22.75	OW test
A11	504	23.30	OW test
A12	504	23.18	OW test
A13	505	0.00	OW test

Extraction: 2 hours
Real-Time PCR : 2 hours
Total time: approx. 1 day

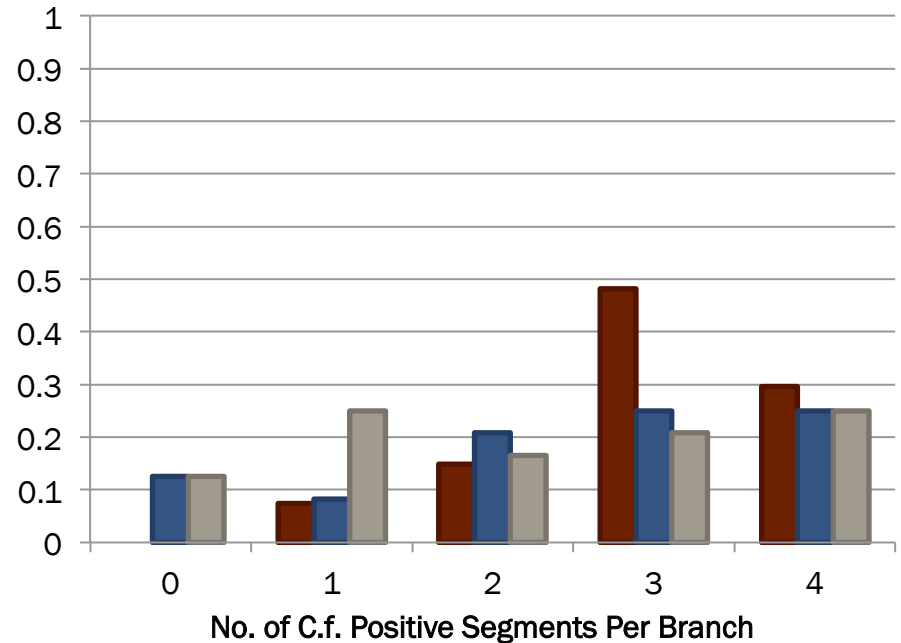
REAL-TIME PCR RESULTS

ACTIVE WILT

Traditional Isolation



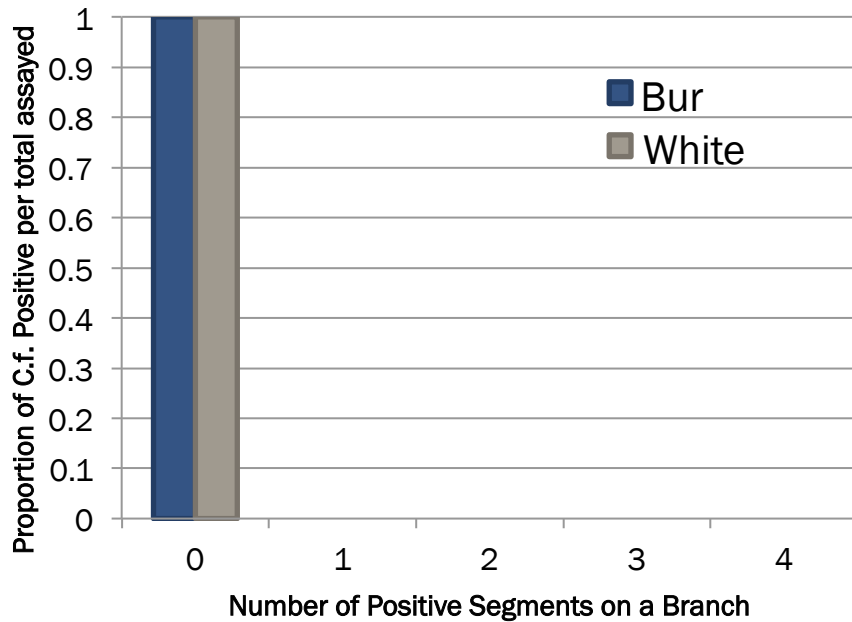
Real-Time PCR



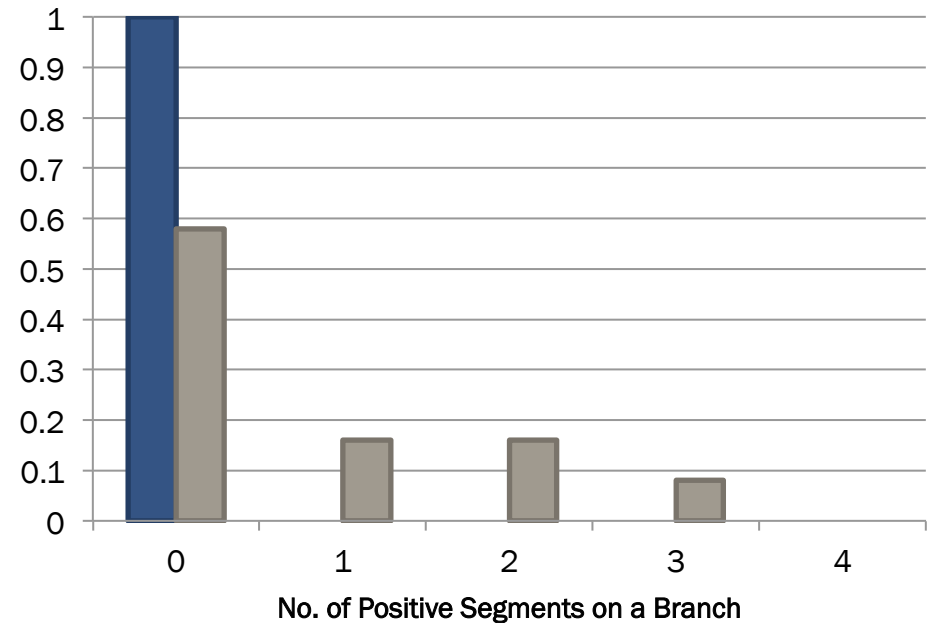
- Red Oak - isolation resulted in more frequent detection
- Bur and White Oak - hard to tell

≥ 1 YEAR DEAD BRANCHES

Traditional Isolation



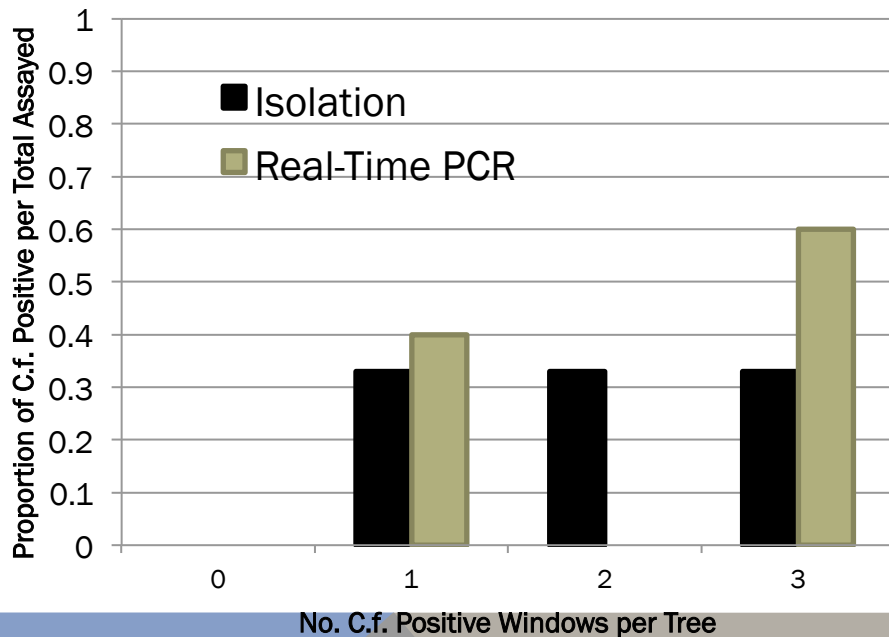
Real-Time PCR



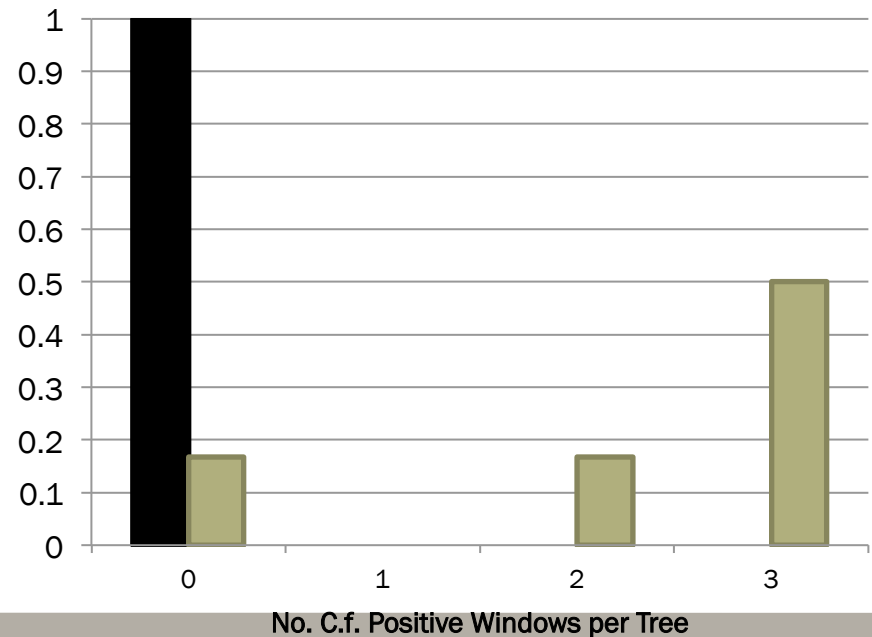
- Bur Oak - Not detected with either
- White Oak - only detected through real-time PCR

RED OAK - MAIN STEM SAMPLES

Streaked Cambium



Mat Scars



- Streaked Cambium - little difference between methods
- Mat Scars - better detection through real-time PCR

NEXT STEPS

Compiling data to compare all three methods to answer questions

Assess usability of new protocols

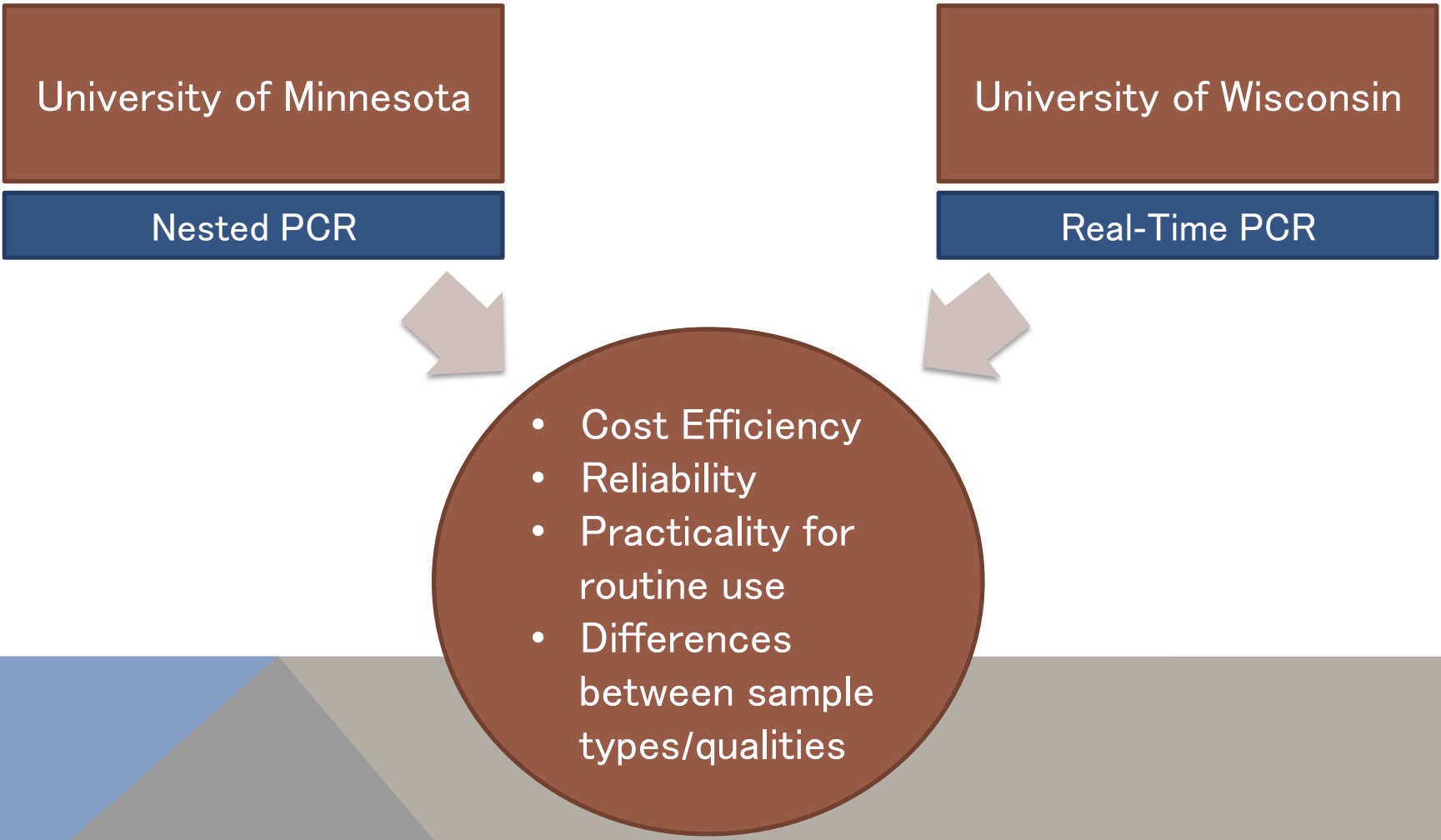
BETA-TESTING NEW PROTOCOLS

University of Minnesota

Nested PCR

University of Wisconsin

Real-Time PCR

- 
- Cost Efficiency
 - Reliability
 - Practicality for routine use
 - Differences between sample types/qualities

TECHNOLOGY TRANSFER

- Work with UMN and UW-Madison to publish in NPDN newsletter
- Other interested diagnostic clinics



ACKNOWLEDGEMENTS

Technical Assistance

- Paul Castillo
- Jameson Scholer
- Kira Ashley

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- Dr. Deborah Samac
- Dr. Gary Johnson
- Dr. Dimitre Mollov

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- Forest Service STDP
- MN Turf and Grounds Foundation
- DOVE Fellowship
- CFANS Fellowship

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Assistance with Study Sites and Trees

- City of Eagan
 - University of Minnesota
 - Three Rivers Park District
 - City of Oakdale
 - Boston Scientific
 - City of Minnetonka
 - Whispering Pines Realty
 - G. Feasky, Private Land Owner
 - City of Apple Valley
- 

THANK YOU!

